Full length research paper

# Evaluation of the biochemical and anti-snake venom effects of *Calliandra Portoricensis* extract fractions in wistar rat models challenged with venom of carpet viper (*Echis Ocellatus*)

Henry P Onyeama<sup>2</sup>; Godwin O Igile<sup>1</sup>; Bob IA Mgbeje<sup>1</sup>; Mbeh U Eteng<sup>1;</sup> Patrick E Ebong<sup>1</sup>

<sup>1</sup>Department of Biochemistry, Faculty of Basic Medical Sciences, University of Calabar, P.M.B 1115, Calabar, Nigeria. <sup>2</sup>National Veterinary Research Institute, Vom, Plateau State, Nigeria.

Accepted 18 March, 2013

Study evaluated effects of flavonoid, polyphenolic and whole ethanolic Calliandra portoricensis leaf fraction on some biochemical indices of venom toxicity in albino Wistar rat. Thirty rats of both sexes were divided into five groups of six rats each. Groups 1 and 2 were normal control (NC) and viperian venom control (VC) respectively. Groups 3, 4 and 5 were venomized with viperian venom and subsequently treated with antidote fractions of flavonoid, polyphenol and whole leaf extracts respectively. Blood samples were analysed for biochemical indices. Phytochemical screening revealed the presence of flavonoids and polyphenols. AST and ALT activity were significantly higher in all treated groups compared to NC. Creatine kinase activity increased significantly in VC compared to NC. These values decreased significantly in fractions/extract treated groups. HGB level decreased significantly while RBC and WBC counts increased significantly in the VC compared to NC. HGB and RBC levels were reversed towards normal by treatment with the fractions/whole extract. Flavonoid fraction only, reversed WBC levels towards normal. Elevated LDL/HDL in VC was decreased towards NC by the plant fractions/extracts. SOD and GPx activity increased significantly in plant fractions/extracts compared to NC and VC. The plant's phytochemical constituents may have significantly lowered lipid peroxidation induced by viperian venom and thus ameliorate hematotoxic effects. This may account for the protection against cardio-toxicity and the shock that normally accompanies viper envenomation.

Key words: Snake venom, envenomation, Calliandra portoricensis, phytochemicals, biochemical indices, wistar rats.

# INTRODUCTION

Deaths due to bites by venomous snakes are common features in the North eastern and central parts of Nigeria. Snake bites in these areas are beginning to constitute a major socio-economic problem, with the ever increasing number of victims of snakebite, due to the favourable breeding and multiplication environment for the snake species (Swaroop and Grab, 1954). Poisonous snakes are a particular problem in Africa and Southeast Asia (Wagstaff et al., 2006) and because not all victims of snakebite get to hospital, only estimates of illness and death are available. One estimate quoted by the World Health Organisation (WHO) is that 2.5 million snake bites occur worldwide each year and 125,000 are fatal (Wagstaff et al., 2006). In West Africa, carpet viper (Echis ocellatus) is the most medically important viperian species (Wagstaff et al., 2006).

Following a viperian envenomation various local tissue alterations occur such as haemorrhage, edema and myonecrosis leading to tissue loss or organ dysfunction (Gutierrez, 1995) and usually these effects develop very rapidly after the snake envenomation, making neutralization by anti-venoms very difficult, especially if serotherapy is delayed due to either late access to

<sup>\*</sup>Corresponding author. Email: patebong@yahoo.com. Tel: +2348037048715

medical care or scarcity of anti-venoms. A known conventional medical procedure involving the use of polyvalent anti-venom (PVA) to neutralize the venom has been in use. However, the use of PVA has many limitations. They are difficult to preserve in rural settings given the erratic nature of power supply; where they are available the PVA is very expensive and beyond the reach of rural people who make up the greater percentage of snakebite victims. Furthermore, life saving anti-venom contains an immunoglobulin pool of unknown antigen specificity and known redundancy which necessitates the delivery of large volumes of heterologous immunoglobulin to the envenomed victim, thus, increasing the risk of anaphylactic reactions, serum sickness and other adverse effects (Wagstaff et al., 2006).Thus the preferred therapy, accessible and affordable, in rural settings is phytotherapy.

Traditional herbalists in south eastern Nigeria have found the leaves and roots of Calliandra portoricensis (Jacq.) Benth., of the family Leguminosae, very useful and effective in neutralizing viperian venom. The extracts of Calliandra portoricensis appear to be specific on the haemotoxic venom of vipers like the rattlesnake, puff adder, and the carpet viper. In distribution, the plant is native to Central America but is abundant in parts of closed forest in Ghana and South eastern part of Nigeria (Irvene, 1961). In traditional preparations used for treating snakebites, the roots are ground to powder or the leaves squeezed in alcohol like wine or alcoholic beverage. It is believed that extracting the secondary metabolites in alcohol enhances the fast absorption of active principles into the blood stream.

Within 15 minutes of entry into the blood stream goose pimples appear on the skin of the snakebite victim, signifying the onset of action of the secondary metabolite and vanish within seconds (Alam et al., 1994; Alam and Gomes, 1998). It is thought that the 'drug' increases the permeability of the capillary membranes as well as that of the gastrointestinal tract (GIT) resulting in the flooding of the GIT with the complex formed by the secondary metabolite and the viperian venom. This results in concomitant stimulation of the emetic centre of the brain. Vomition and diarrhoea ensue with the resultant elimination of the haemotoxic venom from the body. Although, the active ingredient responsible for the antivenom activity of Calliandra portoricensis is not known vet, a phytochemical screening of the plant leaf extract revealed the presence of flavonoids and polyphenols, and more specifically 2-Hydroxy-4-methoxy benzoic acid (Alam et al., 1994; Alam and Gomes, 1998), a secondary metabolite which probably carries out nucleophilic attack on some of the functional groups of the numerous hydrolytic enzymes and non-enzymatic polypeptides involved in carpet viper venom toxicity, thereby neutralizing the venom (Onyeama, 2011). It is also suggested that due to high content of polyphenols in Calliandra portoricensis the neutralization of viperian venom may be due to the complexation between the polyphenols and venom peptides (Houghton,1993). This study therefore is designed to ascertain (i) the phytochemistry and quantify the phytoactive ingredients of *Calliandra portoricensis* (ii) obtain polyphenol and flavonoids-rich solvent fractions and whole ethanolic extract to determine the effects of these extracts on biochemical indices of toxicity and (iii) evaluate the effects of the extracts on phospholipase  $A_2$  enzyme activity in rats challenged with viperian venom.

#### MATERIALS AND METHODS

#### Source of Venom

Lyophilized snake venom of carpet viper was purchased from South African venom suppliers with email address: bewild@worldonline.co.za. The venom sample was preserved as a yellowish crystal inside a tube surrounded by a light-blue layer of silica gel to maintain a dry environment. This was then kept in a desicator at 8°C till it was used.

#### Source of Plant

*Calliandra portoricensis* (Jacq.) Benth. was sourced from the extensive secondary forest of Oji-River in Enugu State where it is used traditionally in the treatment of snakebite of the viperian species. Taxonomically, the plant was identified and confirmed to be *Calliandra portoricensis* by Professor Jonathan C.Okafor, Professor of Botanical Taxonomy, Ebonyi State University, Abakaliki, Nigeria. Voucher specimen "CP-O No.1" has been preserved for reference in the Botany Herbarium of Ebonyi State University, Abakaliki, Ebonyi State, Nigeria.

#### Phytochemical Screening (Analysis)

Harborne's phytochemcial methods (Harborne, 1973) were used to identify alkaloids, flavonoids, saponin, tannins, and polyphenols (Sofowora, 1982). Sulphuric acid spray indicated the presence of glycosides while phlobatanins, anthraquines and hydroxyathraquines were identified by the method of Trease and Evans (1983).

#### Fractionation by Selective Solvent Extraction

The crude methanolic extract of the plant was fractionated by column chromatography using a Glass Column packed with silica gel adsorbent material. A Flavonoid-rich extract fraction was selectively isolated with ethyl acetate and confirmed using the method described by Harborne (1973). Polyphenol fraction was isolated using 30% methanol and confirmed by the Folinciocatean spectrophotometric AOAC technique (AOCS, 1990). The methanol strength was increased to 70% to elute the column to afford a fraction which crystallised to give white needle-like crystalline solids. This was identified as 2-hydroxy, 4-methoxy benzoic acid according to the method described by Alam *et al.* (1994) and Alam and Gomes (1998).

### **Determination of Vitamins and Minerals**

Determination of vitamins C and E was carried out using the method of the Association of Vitamin Chemists as described by Kirk and Sawyer (1989). The versanate EDTA titrametric method (Udoh and Ogunwale, 1986) was employed to determine the calcium and magnesium content of the plant. The heavy metals in the plant were determined using atomic absorption spectrometric technique (AAS) (James,1995).

## Animal treatment

30 albino wistar rats were assigned randomly into five treatment groups of 6 rats each. Group 1 (normal control) received water, the medium for reconstituting the fractions and extracts, intramuscularly. Group 2 (venom control) received 0.2ml of 1mg/ml of viperian venom. Groups 3, 4 and 5 were each injected intramuscularly with 0.2 ml of 1mg/ml (equivalent of 0.2mg) of viperian venom and subsequently injected with 0.5 ml of 100mg/100g body weight (b.w.) anti-dote fractions of flavonoid, polyphenolic cand whole ethanolic extracts of Calliandra portoricensis respectively. This "antidote" was given 4 hours after the administration of the venom. The venom and the extracts were administered intramuscularly. Two hours after the "medication" with plant extracts, the rats were sacrificed after anaesthesia using chloroform vapour. Blood samples were collected from each group via cardiac puncture into sample tubes.

One group of sample tubes contained anticoagulant, Ethylene diamine tetra acetate (EDTA) to prevent blood clotting and allow accurate measurement of haematological parameters. The other group of sample tubes contained whole blood that was allowed to clot and serum separated from them by centrifugation. The sera were then used to determine the lipid oxidation indices. aminotransferases. creatine kinase. phospholipase A<sub>2</sub> enzyme and oxidative stress enzymes activities.

## LD<sub>50</sub> of Plant Extract and Venom

The determination of the LD<sub>50</sub> of plant extract and the method the venom was done using  $LD_{50}$ described by Lorke (1983). The was to determined form of dosage for а basis

subsequent assays employing sub-lethal doses of plant extract and venom.

### **Statistical Analysis**

Analysis of variance (ANOVA) was used in analyzing the data generated by this study. Results were expressed as means  $\pm$  standard deviation. Values of P<0.05, were regarded as being significant.

## RESULTS

Table 1 shows the phytochemical compounds in Calliandra portoricensis, with flavonoids, polyphenols and reducing compounds being the dominant compounds. The mineral elements in the plant are given in table 2 with Ca and Mg being dominant. The result presented in table 3 revealed significant increases (P<0.05) in AST and ALT activities in the venom-treated rats and further significant increases (P<0.05) in the flavonoid-rich, polyphenol-rich and whole CP-extract-treated rats when compared to the control. There was a significant decrease (P<0.05) in CK activity in the flavonoid-rich and polyphenol-rich fractions and a further highly significant decrease (P<0.05) in the whole ethanolic extract when compared to normal control and the venom control groups. The computed AST:ALT ratio (Table 4) for the venom control group showed an increased value compared with the normal control. On the other hand the flavonoid-rich fraction group showed a significant decrease (P>0.05) in AST:ALT ratio, while the polyphenol-rich fraction and whole ethanolic extract groups showed a further statistically significant decrease (P<0.05) in AST:ALT ratio when compared to venom control.

## Table 1 – 4

Table 5 represents venom  $PLA_2$  activity in the serum of experimental rats. The data showed the  $PLA_2$  enzyme activity in the venom control group (0.2104 µmol/min/ml) to be significantly higher (P<0.05) than the  $PLA_2$  activity in the flavonoid-rich, polyphenol-rich and whole ethanolic extract group.

#### Table 5

The anti-oxidant enzymes, superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPx), showed marked decreases (P<0.05) in the venom-treated rats (Figure 1-3)). These oxidative stress (antioxidant) enzymes activities increased significantly (P<0.05) in the plant extracts on the various groups of rats. **Fig 1-3** Assessment of the haematological indices indicated a significant decrease (P<0.05) in the HGB level of venom treated rats, but a significant increase (P<0.05) in the

S/No	Chemical Constituents	Fresh Root	Fresh Leaf	Fresh mixed sample
1	Alkaloids	+	+	++
2	Glycosides	+	+	+
3	Saponins	+	-	+
4	Tannins	-	+	+
5	Flavonoids	++	++	++
6	Reducing compounds	++	++	++
7	Polyphenols	++	+++	+++
8	Phlobatannins	-	-	-
9	Anthraquines	-	-	-
10	Hydoxymethyl anthraquine	+	+	-
	Keys:			

Table 1. Qualitative Constituents of Fresh Calliandra portoricensis.

- + Present
- ++ Present in moderately high level
- +++ Present in high level
  - Absent

 Table 2. Mineral Constituents of C. portoricensis (mg/100g).

Sample	Calcium	Magnesium	Copper	Iron	Selenium
Leaves	30.4	114.3	0.02	0.82	0.02
Stem	218.6	102.6	0.01	0.56	0.02
Root	310.4	121.0	0.02	0.76	0.02

 Table 3. Effects of Treatment on the serum Aspartate Aminotransferase (AST), Alanine Aminotranferase (ALT) and Creatine Kinase (CK) activities on experimental rats (n = 30)

Treatment	AST(u/L) Activity	ALT(u/L) activity	CK(u/L) activity
Control	85.8±4.99	2.47±1.10	651.65±2.92
Venom control	130±0.01	2.68±2.90	693.82±2.54
Flavonoid-rich extract	130±0.01	3.32±1.70	640.70±2.45
Polyphenol-rich extract	141±7.50	9.90±7.40	597.09±6.08
Whole ethanolic extract	140±10.00	18.40±9.90	403.00±110

Enzyme Activities/Ratio					
Control	Venom Control	Flavonoid- rich extract	Polyphenol- rich extract	Whole ethanolic extract	
85.8±4.99	130.0±0	130.0±0	141±7.5	140±0	
2.47±1.1	2.68±2.9	3.32±1.7	9.9±7.4	18.4±9.9	
34.74	48.50	39.16	14.24	7.61	
	85.8±4.99 2.47±1.1	Control         Venom Control           85.8±4.99         130.0±0           2.47±1.1         2.68±2.9	Control         Venom Control         Flavonoid- rich extract           85.8±4.99         130.0±0         130.0±0           2.47±1.1         2.68±2.9         3.32±1.7	Control         Venom Control         Flavonoid- rich extract         Polyphenol- rich extract           85.8±4.99         130.0±0         130.0±0         141±7.5           2.47±1.1         2.68±2.9         3.32±1.7         9.9±7.4	

Table 4. Effects of treatments on the Serum AST: ALT Ratio in experimental rats.

 Table 5. Effects of Treatment on PLA2 Activity in Serum of Experimental Rats (n=30).

Test Group	PLA <sub>2</sub> Activity
Venom control	0.2104µmol/min/ml
Flavonoid-rich extract	0.1195µmol/min/ml
Polyphenol-rich extract	0.0779µmol/min/ml
Whole ethanolic extract	0.0536µmol/min/ml

Flavonoid-rich, polyphenol-rich and whole extract treated rats compared to the venom-treated group (Table 6). The red blood cell counts increased in the venom-treated rats but the CP-extract treated groups maintained red blood cell counts close to the control group suggesting a positive effect on the neutralization of the viperian venom. The administration of venom and the various CP-extracts made no impact on the MCV and MCH. The flavonoid rich and whole extract treated rats showed a significant decrease (P<0.05) on the PCV compared to the venom control and close to normal values.

White Blood Cells (WBC) showed a significant increase (P<0.05) in the venom control (Table 7). This was brought back to near normal value by treatment of the venom-challenged rats with the flavonoid-rich fraction.

The granulocytes, especially the neutrophils, showed a preponderant increase in all the treatment groups when compared to the control. This increase was significant (P<0.05) compared to lymphocyte value which was non-significant (P>0.05) (Table 7).

The values of platelet count (Table 8) showed a significant decrease (P<0.05) probably due to the

overwhelming haemotoxic action of the venom on platelets which the various CP-extracts failed to reverse. Although non-significant changes (P>0.05) were observed in the mean platelet volume (MPV) and platelet distribution width (PDW), the plateletcrit (PCT%) showed significant decrease (P<0.05).

#### Tables 6-8

The effect of treatment on the lipid profile of rats is shown The venom-treated group showed a in Figure 4. significant increase (P<0.05) in total cholesterol level. There was a steady increase in serum triacylglycerols (TG) that was significant (P<0.05) in all groups when compared to the control group. The high density lipoprotein (HDL) content of the venom-treated rats decreased significantly (P<0.05), but increased significantly (P<0.05) in the whole extract treated group The LDL:HDL ratio increased significantly (P<0.05) in the venom-treated rats and decreased significantly (P<0.05) in the whole extract treated group (Table 9). It is believed that the increase in the LDL: HDL ratio which predisposes coronary artery disease may be contributory to the cardiovascular disturbances involved in viperian envenomation.

Blood type		Control	Venom control	Flavonoid-rich extract	Polyphenol-rich extract	Whole Ethanolic Extract		
HGB g/L		177.33±6.81	13.3±8.74	161.±9.72	192.5±6.80	146.6±3.80		
RBC x 10 <sup>12</sup> /L		7.77±0.6	9.21 ±0.43	7.78±0.76	7.96±0.82	7.39±2.58		
HCT% (PCV)		44.7±3.2	52.45±0.94	46.11±3.4	52.4±1.93	46.94±4.71		
MCV (fL)		57.75±2.7	54.72±1.36	57.6±2.9	58.4±0.53	57.96±3.2		
MCH (pg)		17.12±0.48	17.4±0.33	18.1±0.46	17.82±0.63	17.74±1.3		
MCHC g/L		297±6.84	312±10.91	313.67±9.11	310.3±10.4	317.2±6.05		
HGB	=	Haemogle	Haemoglobin concentration					
RBC	=	Red Bloo	d Cell					
HCT	=	Haemato	crit, MCV = Me	ean corpuscular v	volume			
MCH	=	Mean Cell Haemoglobin						
			Ū					
MCHC	=	Mean Cell Haemoglobin concentration.						
Pg	=	(Pico grar	(Pico gram or micro gram)					
5			5	,				

 Table 6. Effects of Treatment on Red Blood Cells of Experimental Rats (n=30).

Table 7. Effects of Treatment on White Blood Cells of Experimental Rats (n=30).

Blood type	Control	Venom Control	Flavonoid-rich extract	Polyphenol- rich extract	Whole ethanolic extract
WBC x 10 <sup>9</sup> /L	$10.3\pm1.94$	14.1±5.08	10.5±4.21	15 ±2.49	13.86±2.3
Lymph x10 <sup>9</sup> /L	5.3±1.0	4.15±1.39	4.22±2.13	3.83±1.07	4.26±2.61
Gran x 10 <sup>9</sup> /L	3.4±1.7	7.52±4.72	5.5±2.85	15.2±6.2	8.82±4.5
Lymph%	52.03±3.17	29.9±4.20	38.9±4.50	19.6±4.83	29.54±4.69
Gran %	38.35±4.86	61.9±4.08	55.05±5.05	74.35±5.28	64.54±4.69

WBC = White Blood Cells, Lymph = Lymphocyte, Gran = Granulocyte

# Fig 4

### Table 9

## DISCUSSION

The outcome of the evaluation of the biochemical and anti-snake venom effects of *Calliandra portoricensis* 

extracts revealed increases in AST and ALT in the various treatment groups (Table 3). The computed AST:ALT ratio (Table 4) gave a significant value (P<0.05) in the serum of rats treated with the venom. The serum of rats treated with the various plant extract fractions showed a non-significant value (P>0.05) of AST:ALT ratio (Table 4). Although results on the effect of viperian venom administration on aminotranferase activities on tissues have not been reported in literature, the use of

Blood type	Control	Venom control	Flavonoid- rich extract	Polyphenol- rich extract	Whole ethanolic extract
PLT x 10 <sup>9</sup> /L	551 ±1.70	416.67±1.32	237.8±1.96	205±1.33	111.6±0.87
MPV(fL)	8.57±0.44	9.12±0.44	10.6±0.55	9.1±0.78	9.86±0.55
PDW	14.67±0.23	14.77±0.23	14.82±0.67	14.7±0.55	14.92±0.19
PCT %	0.47±0.13	0.37±0.10	0.235±0.16	0.269±0.09	0.106±0.09

Table 8. Effects of Treatment on platelets of Experimental Rats (n=30).

PLT =	Platelets
MPV=	Mean Platelets Volume
PDW	= Platelets Distribution Width
PCT%	= Plateletcrit

Table 9. Effects of Treatment on LDL:HDL Ratio in Experimental Rats.

Test Serum Lipid	Control	Venom Control	Flavonoid rich extract	Polyphenol- rich extract	Whole Ethanolic extract
LDL	64.33±1.33	105.78±5.91	31.96±0.26	12.25±2.79	61.17±1.23
HDL	44.74±1.03	11.17±5.70	72.24±1.16	107.39±1.59	165.13±1.13
LDL: HDL	1.44	9.47	0.44	0.11	0.37

AST:ALT ratio may generally be used as a biomarker to monitor pathologies implicating the heart or liver as has been reported elsewhere (Howcroft,1987). According to these reports, an increase in AST:ALT ratio points to a pathology involving the heart while the reverse implicates the liver (Howcroft,1987). The increase in this ratio observed in the venom-treated animals therefore provided experimentally determined evidence, at molecular level, that viperian venom toxicity was indeed cardiovascular (Table 4).

Thus the increase in AST and ALT activities and the decrease in AST:ALT ratio pointed to increase in the detoxification ability and capacity of the liver to handle the venom-extracts complex which brought about increase in the ALT activity and the consequent decrease in the AST:ALT ratio in rats treated with the various CP-extract fractions (Table 4).

Phospholipase A<sub>2</sub> enzyme (PLA<sub>2</sub>) appears to be the lethal component of viperian venom (Onyeama *et al.*, 2012).

There were significant decreases in PLA<sub>2</sub> activity (p<0.05) in the CP-extract groups (Table 5). This decrease in PLA<sub>2</sub> activity may be attributed to the complexation of the venom PLA<sub>2</sub> with the polyphenolics and the reduction in the free radicals as a result of the flavonoid component of the plant extract (Onyeama et al. 2012). As a result of the complexation of the  $PLA_2$  with the polyphenolics (Houghton, 1993), there was a diminished PLA<sub>2</sub> activity and the subsequent inactivation of the venom by the CP-extract fractions. It can thus be inferred that these extracts neutralized PLA<sub>2</sub> enzyme activity which elicits all the damages and changes at molecular level, once there is viperian venom in the blood stream. The anti-oxidant activities in the groups showed sera of CP-extracts treated statistically significant increases (P<0.05). In this study, viperian venom appeared to have obliterated or overwhelmed the naturally-existing antioxidant enzymes activities in rats.

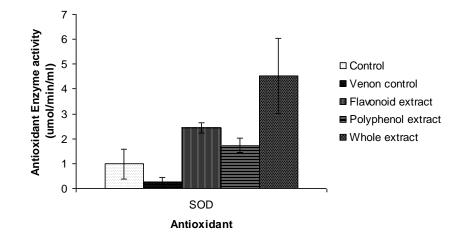


Figure 1. Effects of Extract fractions on Serum Anti-oxidant Enzyme (SOD) Activity.

Figure 2. Effects of Extract fractions on Serum Anti-oxidant Enzyme (CAT) Activity.

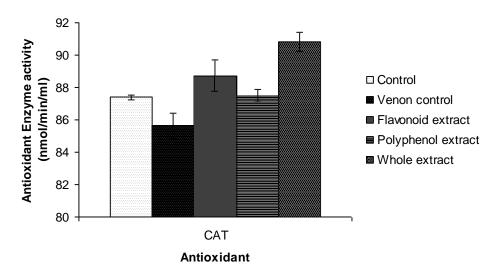


Figure 3. Effects of Extract fractions on Serum Anti-oxidant Enzyme (GPx) Activity.

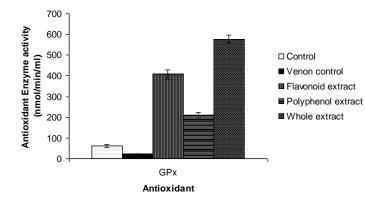
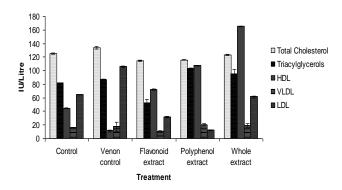


Figure 4. Effects of Extract fractions on Serum Lipids and Cholesterol



This appeared much more marked on SOD (Figure 1) activity and consequently on CAT (Figure 2) and GPx (Figure 3) indirectly. The increases in the CP-extract groups could be attributed to the heavy quantitative content of flavonoid-rich extract, the complexation of polyphenolics with  $PLA_2$  (Houghton,1993) which is responsible for the generation of massive amounts of free radicals in viperian envenomation. The complexation of the polyphenols with Ca<sup>2+</sup>-dependent  $PLA_2$  and the marked anti-oxidant activity of the flavonoid content of *Calliandra portoricensis* seemed to have worked synergistically and in tandem with one another to enhance overall, the anti-oxidant potentials of the plant extract.

Haematological indices suggested a normalization of the blood parameters after treatment with CP whole extracts and fractions (Table 6 and 7). The spectrum of changes in lipid parameters especially the elevation in HDL (Figure 4) on the one hand and the rapid return of RBCs and WBCs to near normal values on the other have shown that Calliandra portoricensis extracts had a neutralizing effect on the venom. The decrease in LDL:HDL ratio ((Table 9) proves that Calliandra portoricensis extracts removed the possibility of artherogenesis which the venom can predispose a victim to after recovery. In fact epidemiological studies indicate that high plasma HDL levels are strongly correlated with a low incidence of cardiovascular complication (Voet and Voet,1990).

## CONCLUSION

Exposure of albino wistar rats to viperian venom negatively, their enzyme, lipid and alters haematological parameters. The venom is both cardiotoxic and haemotoxic and may bring life to an acute fatality if not treated immediately. The alteration of these biochemical parameters accounts for the viperian venom toxicity. Phytochemical fractions of flavonoids. and polyphenolics whole extract of Calliandra portoricensis have been able to reverse these altered biochemical changes to a very significant (P<0.05) extent. The effect of CP-extract fractions especially in lowering serum lipid level in envenomed rats with elevated plasma lipid levels, the marked PLA<sub>2</sub> enzyme activity decrease in and the reduction in free radicals as a result of the flavonoid component of the plant extract may be attributed to the complexation of the venom PLA<sub>2</sub> the anti-oxidative secondary compounds of by Calliandra portoricensis. Thus, we can conclude that this anti-venom property of Calliandra portoricensis exploited for therapeutic can be benefits.

# REFERENCES

Alam MI, Gomes A (1998). Adjuvant effects of antiserum action potentiation by a (herbal) compound 2-hydroxy, 4methoxy benzoic acid isolated from root extract of the Indian medicinal plant 'Sarsasparilla' (*Hemidesmus indicus*). Toxicon, 36: 1423-1431.

Alam MI, Auddy B, Gomes A (1994). Isolation, purification and partial characterisation of viper venom inhibiting factor of root extract of Indian medicinal plant Sarsaparilla *(Hemidesmus indicus,* R. Br.). Toxicon, 32: 1551-1557.

AOAC (1990). Official Method of Analysis of the Association of Analytical Chemists. *AOAC*. Washington D. C., pp 223-225.

Gutierrez JM (1995). Clinical Toxicology of Snakebite in Central America. In: Meier. J. White (Ed). *Handbook of clinical toxicology of animal venoms and poison*. CRC Press ,Florida, pp 645 -665.

Harborne JB (1973). Phytochemical Methods. *A guide to Modern Technique of Plant Analysis.* Chapman and Hall, London.

Houghton PJ (1993). In vitro testing of some West African and Indian plants used to treat snakebites. *Medicament et Nutrition: L' Approche ethnopharmacologique*, 263– 274.

Howcroft D (1987). Diagnostic enzymology: Analytical Chemistry by open Learning ( $1^{st}$  Edition) James, A. M (Ed). John Willey and Sons, New York, pp 156 – 210.

Irvene FR (1961). *Woody Plants of Ghana*. Oxford University press, London, p 869.

James CS (1995). *Experimental Methods in Analytical Chemistry of Foods*. Chapman and Hall. New York, pp 28.

Kirk RS, Sawyer R (1989). Pearson's Food Composition and Analysis. Longman, London.

Lorke D (1983). A new approach to practical Acute Toxicity Test. *Institute Fur Toxicologie*, Bayer A.G, Friedrich – Ebert- Strasse. 217, D- 5600 Wuppertal, Federal Republic of Germany.

Onyeama HP (2011). Evaluation of the Biochemical and Anti-Snake venom effects of *Calliandra portoricensis* extract in Wistar Rats challenged with venom of Carpet viper (*Echis oscellatus*) (PhD Thesis, University of Calabar).

Onyeama HP, Ebong PE, Eteng MU, Igile GO, Ibekwe HA, Ofemile PV (2012). Histopathological Responses of the Heart, Liver and Kidney to *Calliandra Portoricensis* Extracts in Wistar Rats Challenged with Venom of *Echis Ocellatus.* J. Applied Pharm. Science. 6(2): 164-177.

Sofowora EA (1982). Medicinal Plants and Traditional Medicine in Africa. John Wiley and Sons, New York.

Swaroop S, Grab B (1954). Snakebite mortality in the World. Bull *WHO*, *61*, 949 – 956.

Trease GE, Evans EC (1983). Pharmacology (12<sup>th</sup> ed.). Bailliere Tindall, London.

Udoh JE, Ogunwale JA (1986). Laboratory Manual for the Determination of Soil, Plant and Water Samples. University Press, Ibadan.

University Press, Ibadan. Voet D, Voet JG (1990) *Biochemistry* (1<sup>st</sup> ed.). John Wiley and Sons, New York, pp. 557, 773 – 1159. Wagstaff SC, Laing GD, Theakston DG, Papaspyridis C, Harrison RA (2006). Bioinformatics and Multiepitope DNA Immunization to Design Rational Snake antivenom. Plos Medicine, 3 (6):184.