

International Journal of Agricultural Sciences ISSN: 2167-0447 Vol. 9 (7), pp. 001-005, July, 2019. Available online at www.internationalscholarsjournals.org © International Scholars Journals

Author(s) retain the copyright of this article.

Full Length Research Paper

Mineralization of nitrogen from ¹⁵N labeled crop residues at varying temperature and clay content

M. De. Roy^{1,2*}, P. K. Chhonkar¹ and A. Patra¹

¹Division of Soil Science and Agricultural Chemistry, Indian Agricultural Research Institute, New Delhi–12, India. ²STL, Pulses and Oilseed Research Station, Berhampore, West Bengal.

Accepted 24 March, 2019

A laboratory incubation experiment was carried out for 85 days to monitor the N mineralization during the decomposition of ¹⁵N labeled wheat and rice straws incorporated at 5 g kg⁻¹ soil at varying temperatures and with soils of varying clay content. Results have shown that N mineralization surpass immobilization 40th day onwards. Irrespective of temperature and clay content, mineralization of rice residue N was found to be significantly higher than that of wheat residue throughout the incubation period. Increasing temperature and clay content both had positive effect on residue N mineralization. At 85th day after incubation N mineralization from the residues was found to be 13 - 14%. Clay content, temperature, composition of the residues are important factors controlling plant residues decomposition under laboratory conditions.

Key words: Rice and wheat residues, straw, N-mineralization, immobilization.

INTRODUCTION

Rising costs of chemical fertilizers, as well as their possible long term adverse effects on soil health, crop productivity and environmental pollution have resulted to more attention on the recycling of organic wastes. Incorporation of organic materials through crop residues may be an important choice for maintaining or increasing soil fertility as it is well recognized that the organic matter is a key attribute of soil fertility. Crop residues have a great potential in India particularly in predominantly rice, wheat and sugarcane growing areas. The traditional practice of burning eliminates 70 to 80% of the N held in the crop residue (Sarkar et al., 2000). Consequently, the change from burning to straw incorporation will likely facilitate the cycling of N in residues back to the soil. Crop residues undergo mineralization before N becomes available to crops. Availability of crop residue nitrogen to succeeding crops is determined by the outcome of mineralization-immobilization processes (Paul and Juma, 1981; Jansson and Persson, 1982). Powlson et al. (1995) stated that net immobilization of soil nitrogen generally occurs when residues have a high C: N (>25:1). Soil temperature and texture as well as composition of straws

are believed to exert a significant influence on N mineralization-immobilization dynamics of organic residues either through changing soil biota, enzymatic activities or through adsorption of soil organics (Singh et al., 1991). Except few works (Patil and Sarkar, 1993), not much information is available regarding the effect of crop residues incorporation on N dynamics at varying soil temperature and texture.

Therefore, the present laboratory study for 85 days was carried out to generate comprehensive informations regarding the effects of soil incorporation of rice and wheat straws on the dynamics of residue N mineralization-immobilization at varying temperatures and clay content, and to examine the residue inorganic N content in terms of NH_4^+ and NO_3^- at various intervals after incubation. ¹⁵N stable isotope methodology was used to follow the turnover of labeled crop residue N in the inorganic pools.

MATERIALS AND METHODS

Laboratory experiment using soil with varying clay content and temperature was carried out for 85 days at the Division of Soil Science and Agricultural Chemistry, IARI, New Delhi, India. Labeled crop residues (rice and wheat straws) obtained from a previous greenhouse experiment using 20% atom excess urea were used.

^{*}Corresponding author. E-mail: mithuderoy@yahoo.in.

Crop residue (C)							
	0	10	20	40	60	85	Mean
Control	19.1	21.1	23.4	25.5	26.8	27.1	24.8
Wheat	19.1	8.5	10.9	26.9	30.4	32.0	21.7
Rice	19.1	8.8	13.6	28.6	30.9	31.5	22.6

Table 1. Effect of types and levels of crop residues on inorganic nitrogen ($NH_4^+ + NO_3^-$) content mg kg⁻¹ at different days of incubation.

Sources: CD at (P = 0.05); C = 0.60. Data represented in this table are the mean values averaged over three clay contents and two temperatures.

Table 2. Effect of clay content of soil on inorganic nitrogen $(NH_4^+ + NO_3)$ content mg kg⁻¹ at different days of incubation.

Soil of varying clay								
content (S)	10	20	40	60	85	wean		
So	14.8	17.7	29.7	32.1	33.8	25.7		
S1	13.4	16.5	29.1	31.1	31.5	23.3		
S ₂	11.6	14.3	22.9	25.4	26.1	20.1		

Sources: CD at (P = 0.05); S = 0.60. S₀, S₁, S₂ refer to original soil, 75% original soil + 25% sand and 50% original soil + 50% sand respectively and data represented in this table are averaged over three residue levels and two temperatures.

Table 3. Effect of temperature and days of incubation on inorganic nitrogen ($NH_4^+ + NO_3^-$) mg kg⁻¹ content of soil.

	Days of incubation (D)								
Temperature (T)	0	10	20	40	60	85	Mean		
T 1	19.1	12.5	16.1	26.7	28.9	29.8	22.7		
T ₂	19.1	13.2	15.8	27.3	29.8	30.6	23.4		

Sources: CD at (P = 0.05); S = 0.49. T₁, T₂ refer to 20 and 35° C, respectively and data represented in this table are averaged over all three residue and clay contents levels.

The initial C:N ratios of rice and wheat straws were 82.9:1 and 89.5:1, respectively. The bulk surface soil (0 - 15 cm) of loamy texture belonging to Typic Haplustept (Inceptisol) collected from the Institute Farm had the following characteristics; clay 17.7%; pH 8.3, organic C 3800 mg kg⁻¹, total N 680 mg kg⁻¹, NH_4^+ - N 10.2 mg kg⁻¹, NO_3 -N 15.3 mg kg⁻¹, field capacity (% moisture at 1/3 bar) 18.20. To get the soil with different clay contents (17.7, 13.3 and 8.85%), it was sieved (2 mm) and proportionately mixed with previously washed Jamuna sand at 0, 25 and 50%, respectively. Sterilized (autoclaved) white plastic pots were filled with 1 kg of these soils. Crop residues (viz., no straw, wheat straw and rice straw) were chopped to 2 mm pieces and mixed at 5 g kg⁻¹ soil to the respective pots. Two sets of pots were placed in the rack of respective BOD incubators at a constant temperature of 20 and 35°C. Moisture content of the pots was maintained at field capacity (FC) throughout the incubation period by measuring the difference in weight loss. Soil samples were collected at 10, 20, 40, 60 and 85 days after incorporation of crop residues and analyzed for NH4⁺-N, NO3⁻N and ¹⁵N. NH₄⁺-N and NO₃⁻ -N were determined by steam distillation using the procedure described by Bremner and Edward (1965) and Bremner and Keeney (1966), respectively. Subsequently the titrated distillate was evaporated at 45°C for ¹⁵N:¹⁴N ratio analysis using Duma's method (Keeny and Nelson, 1982). The experiment was conducted in a factorial randomized complete block with 3 replications and statistical analysis of the data was performed

applying SPSS software package and LSD was used for treatment means comparison as per the methods suggested by Gomez and Gomez (1984).

RESULTS AND DISCUSSION

Incorporation of wheat and rice residue significantly reduced the inorganic N content from 19.1 mg kg⁻¹ under unamended soil to 8.5 and 8.8 mg kg⁻¹ soil for wheat and rice residue treated soils respectively within 10 days of incubation. From 10th to 20th day, no net N mineralization was observed but inorganic N level increased significantly from 40th to 85th day over control (Table 1). Higher clay content and temperatures have shown more inorganic nitrogen level and irrespective of levels of clay content and temperatures, inorganic nitrogen increased significantly from 10 days onwards throughout the incubation period (Tables 2 and 3). Maximum mean inorganic N content for wheat and rice residue treatments were recorded as 32.0 and 31.5 mg kg⁻¹ respectively at 85th day. But the value was found statistically at par



Figure 1. Mineralization of Nitrogen from ¹⁵N labeled crop residues under varying clay content. S₀, S₁, S₂ refer to original soil, 75% original soil + 25% sand and 50% original soil + 50% sand respectively.

par for the residue treatments. However, rice residue treatment exhibited significantly higher level of mean inorganic N content than wheat residue treat-ment during 20th and 40th day (Table 1). Increasing the temperature from 20 to 35°C increased inorganic N content from 12.5 to 13.2 mg kg⁻¹ at 10th day, 26.7 to 27.3 mg kg⁻¹ at 40th day, and 29.8 to 30.6 mg kg⁻¹ at 85th day of incubation (Table 3). N-immobilization in terms of NH_4^+ - N + NO₃- N estimated over control following straw incorporation was found to be prominent up to 20th day, thereafter no net mineralization was found up to 40th day. However, after 40th day, N mineralization exceeds that of immobilization ranging from 14 - 17% at 60th day and 19 - 20% at 85th day as estimated over control that is, by the difference method (Table 1). Wagger et al. (1985) demonstrated mineralization of N from ¹⁵N labeled wheat and sorghum residue to be 12.5 - 14.7% and 14.9 - 32.9% respectively during one crop season. More or less similar results were also reported by Sarmah and Bordoloi (1994) who found conspicuous net immo-bilization of N up to 4 weeks in soils due to treatment with wheat straw after which mineralization exceeded immo-bilization. However, Christensen (1985) observed net immobilization just after wheat and barley straw burial in a coarse and sandy loam soil under field condition reaching maximum after six months. This may be due to the fact that mineralization rates in field soils used to be lower than those reported in controlled environment since conditions are not continuously favourable for biological activity in natural ecosystems. Higher clay content in the original soil as compared to sand mixed soils caused more retention of inorganic N either due to the presence of more aerobic microsites followed by less loss through denitrification or more stabilization of the microbes by the formation of the so-called 'clay envelop' which becomes effective

against predation (Marshall, 1975). Positive effect of clay on mineralization has been found by Campbell and Sauster (1982). However, negative effects were also reported by Van Veen et al. (1985) and Amato et al. (1987). Increasing the temperature from 20 to 35°C resulted in higher inorganic N content in the incubated soil irrespective of residues or clay content. N mineralization is a temperature dependent microbial process with optimum temperature being 35°C (Poovarodam and Tatelll, 1988) or 30 to 35°C (Cassman and Munns, 1980).

The rate of mineralization of N from labelled crop residues in soil was considerably higher at higher clay content and higher temperature (Figures 1 and 2). Per cent inorganic N recovered in soil from labeled wheat and rice residues were 5.69 and 6.47% at 20th day, 7.61 and 8.53% at 40th day and 10.03 and 10.76% at 60th day and 13.22 and 13.81% at 85th day after incubation, respectively (Table 4). Significant decrease in mean inorganic N derived from crop residues was recorded for decreasing clay content. Moreover, temperature had positive effect on residue mineralization showing higher mean inorganic N (8.97%) at 35 than 20°C (7.39%). At 85th day of incubation per cent inorganic N recovered in soil from labeled wheat and rice residue was 13.22 and 13.81%, respectively (Table 4). Comparatively higher value (19 - 20%) obtained from difference method (Table 1) may be due to the enhancement of soil organic N mineralization on incorporation of crop residues. The present data are in agreement with the findings of Amato et al. (1987). Similarly Wagger et al. (1985) demonstrated mineralization of N from ¹⁵N labeled wheat and sorghum residues to be 12.5 - 14.7% and 14.9 - 32.9%, respectively during one crop season. Mineralization of rice residue was found to be significantly higher than the wheat residue. This phenomenon may be due to higher



Figure 2. Mineralization of nitrogen from ¹⁵N labeled crop residues under varying temperatures. T₁ and T₂ refer to 20 and 35°C, respectively.

Table 4. Per cent inorganic N (NH ₄ ⁺ + NO ₃ ⁻) recovered in soil from	n ¹³ N labeled crop residues under varying temperatures and clay
content of soils at different days of incubation.	

4 -

•	Soil of varying clay	Days of incubation (D)							
rosiduos		20		40		60		85	
residues	content	T 1	T2	T1	T2	T 1	T2	T1	T2
Wheat	So	6.32	6.28	8.13	8.65	10.99	11.52	14.45	14.81
	S ₁	5.64	5.82	7.32	7.76	9.98	10.25	13.08	13.67
	S ₂	5.01	5.16	6.64	7.21	8.56	8.84	11.24	12.01
	Mean	5.69		7.61		10.03		13.22	
Rice	So	7.01	7.22	9.14	10.19	11.56	12.62	14.02	15.85
	S ₁	6.11	6.92	7.65	8.47	10.29	11.44	13.34	14.15
	S ₂	5.69	5.92	7.81	7.95	8.94	9.71	11.55	13.95
	Mean	6.47		8.53		10.76		13.81	
Sources		CD at (P = 0.05)							
С		0.30							
S		0.37							
Т						0.30			

NB : S₀, S₁, S₂ refer to original soil, 75% original soil + 25% sand and 50% original soil + 50% sand respectively and T₁, T₂ refer to 20^{0} C and 35^{0} C, respectively

lignin content of wheat residue over rice residue as the C:N ratio was more or less similar for both the residues. The nature and content of lignin is a good index for prediction of net N release (Berg and McClaughterty, 1987). In a study with several parameters Muller et al. (1988) concluded that lignin content was better than N concentration or C:N ratio in predicting the amount of mineralized N. Increasing temperature from 20 to 35°C increased the per cent inorganic N derived from residue. This may be due to higher degree of mineralization at higher temperature. Similarly increasing clay content also showed positive effect on residue N mineralization. These findings are in agreement with the observations of Patil and Sarkar (1993). Higher temperatures and soil clay content led to best N mineralization from residues in the present study as evidenced from the Table 4. Thus, it can be concluded that amount of clay in soil, temperature and residue type play an important role in modifying the mineralization and immobilization pattern of crop residue nitrogen. Therefore, nature of soil, temperature, residue type and period of incorporation should be assessed to get maximum benefit of available N pools during straw decomposition.

REFERENCES

- Amato H, Ladd JN, Ellington A., Ford G, Mohaney JE, Taylor A, Walsgatt D (1987). Decomposition of plant material in Australian soils. IV. Decomposition in situ of 14C and 15N labeled legume and wheat materials in a range of southern Australian soils. Australian J. Soil Res., 25: 95-105.
- Berg B, McClaughterty C (1987). Nitrogen release from litter in relation to the disappearance of lignin. Biochemistry, 4: 219-224.
- Bremner JM, Edward AP (1965). Determination and isotope ratio analysis of different forms of nitrogen in soils.I. Apparatus and procedure for distillation of ammonium. Soil Sci. Soc. Am. Proc., 29: 504-507.
- Bremner JM, Keeney DR (1966). Determination and isotope ratio analysis of different forms of nitrogen in soils. III. Exchangeable ammonium, nitrate and nitrite by extraction – distribution methods. Soil Sci. Soc. Am. Proc., 36: 577-582.
- Campbell CA, Sauster W (1982). Loss of organic matter and potentially mineralized nitrogen from Saskatchewan soils due to cropping. Can. J. Soil Sci., 62: 651-656.
- Cassman KG, Munns DN (1980). Nitrogen mineralization as affected by soil moisture, temperature and depth. Soil Sci. Soc. Am. J., 44: 1233-1237.
- Christensen BT (1985) . Wheat and barley straw decomposition under field condition. Soil Biol. Biochem., 17: 691-697.
- Gomez AK, Gomez AA (1984). Statistical procedures for agricultural research. 2nd edition. John Wiley and Sons. New york.
- Jansson SL, Persson J (1982). Mineralization and immobilization of soil nitrogen. In : Nitrogen in Agricultural Soils (F.J. Stevenson, Ed.,), Am. Soc. Agron., Madison, pp. 229-252.

- Keeney DR, Nelson DW (1982). Nitrogen Inorganic forms. In: Methodes of soil analysis. A.L. Page el al (ed.). Part 2. 2nd ed. Agron Monogr. ASA and SSSA, Madison, W.I. pp. 643-698.
- Marshall KC (1975). Clay mineralogy in relation to survival of soil bacteria. Annu. Rev. Phytopathol., 13: 127-134.
- Muller MM, Sundman V, Soininvaara O, Merilainen A (1988). Effect of chemical composition on the release of N from agricultural plant material decomposing in soil under field condition. Biol. Fert. Soils, 6: 78-83.
- Patil RG, Sarkar MC (1993). Mineralization-immobilization of nitrogen in soils mixed with wheat straw. J. Indian Soc. Soil Sci., 41(1): 33-37.
- Paul EA, Juma NG (1981). Mineralization and immobilization of soil nitrogen by microorganism. In F.E. Clark and T. Rosswall (ed.) Terrestrial nitrogen cycles. Ecol. Bull., 33:179-195.
- Poovarodam S, Tate III RL (1988). Nitrogen mineralization rate of acidic Xeric soils of New Jersey Pine lands : laboratory studies. Soil Sci., 145(5): 337-344.
- Powlson DS, Jenkinson DS, Pruden G, Johnston AE (1985). The effect of straw incorporation on the uptake of nitrogen by winter wheat. J. Sci. Food Agric., 36: 26-30.
- Sarkar A, Gangwar B, Yadav RL, Bhatia PC (2000). Crop residue A potential natural resource of plant nutrients. International conference on managing natural resources for sustainable agricultural production in the 21st century. Feb, 14-18, 2000, I.A.R.I, New Delhi, 1089-1090 pp.
- Sarmah AC, Bordoloi PK (1994). Decomposition of organic matter in soils in relation to mineralization of carbon and nutrient availability. J. Indian Soc. Soil Sci., 42(2): 199-203
- Singh Y, Khind CS, Singh B (1991). Efficient management of leguminous green manures in wet land rice. Adv. Agron., 45: 135-189.
- Van Veen J, Ladd JN, Amato H (1985). Turnover of C & N through microbial biomass in a sandy loam and a clay soil incubated with [14C(V)] glucose and (¹⁵N) (NH₄)₂ SO₄ under different moisture regimes. Soil Biol. Biochem., 17: 747-756.
- Wagger MG, Kissel DE, Smith SJ (1985). Mineralization of nitrogen from 15N labelled crop residues under field conditions. Soil Sci. Soc. Am. J., 49: 1220-1231.