Full Length Research Paper

Studies on the impact of water deficit on morphological, physiological and yield of banana (*Musa spp*.,) cultivars and hybrids

K. Krishna Surendar^{*1}, D. Durga Devi¹, I. Ravi², P. Jeyakumar¹ and K. Velayudham³

^{*1}Department of Crop Physiology, TNAU, Coimbatore-641 003, India. ²National Research Centre for Banana (ICAR), Thiruchirapalli, India. ³Department of Farm Management, TNAU, Coimbatore-641 003, India.

Accepted 27 February, 2013

Field investigations were undertaken to screen the banana cultivars and hybrids for water stress tolerance and to elucidate information on morphological, physiological and yield characteristics mechanism of banana cultivars and hybrids. Stress was imposed at different critical stages viz., 3rd, 5th, 7th and 9th month after planting. The stress was given by scheduling irrigation at the 50 per cent available soil moisture (ASM) characteristic during critical stages. The soil moisture content was analyzed by using pressure plate membrane apparatus. In control plots, the irrigation was given at the ASM of 80 per cent with the soil water potential of around -6 bars and in the case of stressed plots; the irrigation was given when an ASM reached 50 per cent with the soil water potential of -14 bars. In stressed plots, 50 per cent ASM was reached around 30 days. In this present study conducted with twelve cultivars and hybrids with three replications. The data were analyzed by using split plot design. The morphological characters viz., plant height, and number of leaves were significantly enhanced by control when compared to stress-treated plants and physiological characters like RWC, total chlorophyll content and osmotic potential can be considered good indicators of leaf water status in banana and can therefore be used for irrigation scheduling. Among the twelve cultivars and hybrids, Karpuravalli, Karpuravalli x Pisang Jajee, Saba, and Sannachenkathali was identified as tolerant to water stress and showed lesser reduction in the range of 3 to 14 per cent in morphological characters and 8-10 per cent over control in physiological characters leads to maintained its superiority over control and get higher bunch yield; whereas, Matti, Pisang Jajee x Matti, Matti x Anaikomban and Anaikomban x Pisang Jajee were notified as sensitive cultivars and hybrids with mean reduction of 22 per cent in morphological and physiological characters than control due to irrigation at 50 ASM.

Key words: Water deficit, morphological, physiological characters, yield, banana cultivars.

INTRODUCTION

Banana is the 'queen of tropical fruits' and is one of the oldest fruits known to mankind from pre-historic times. Today, it is the leading tropical fruit in the world market with a highly organized and developed industry. It is the fourth largest fruit crop in the world after grapes, citrus fruits and apples. Drought is an insidious hazard of nature. Although it has scores of definitions, it originates from a deficiency of precipitation over an extended period of time, usually a season or more. This deficiency results in a water shortage for some activity, group, or environmental sector. Water deficit occurs when water potentials in the rhizosphere are sufficiently negative to reduce water availability to sub-optimal levels for plant growth and development. On a global basis, it is a major cause limiting productivity of agricultural systems and food production (Bray et al., 2000). Banana plant productivity is greatly affected by environmental stresses such as drought, water and cold. Plants respond and adopt to these stresses to survive under stress condition at the molecular and cellular levels as well as at the physiological and biochemical levels. Physiological responses to soil water deficit are the feature that is most

^{*}Corresponding author. E-mail: surendartnau@gmail.com



7th MAP

-\$4

-S10

\$3

59

M₁ - Control, M₂ - Stress

Figure 1. Effect of water stress on Relative Water Content (%) of banana cultivars at different growth stages.

S₁- Karpuravalli S₂- Karpuravalli x Pisang Jajee S₃ - Saba S₄ - Sannachenkathali S₅ – Poovan S₆ – Neypoovan

3rd MAP

-S1

-\$7

5th MAP

S2

\$8

0.0

S₇ – Anaikomban S₈ - Matti x Cultivar Rose S₉ - Matti S₁₀ - Pisang Jajee x Matti S₁₁ - Matti x Anaikomban S₁₂-Anaikomban x Pisang Jajee

Harvest

-S12

9th MAP

-85

-S11

likely to determine the response of the crop to irrigation. The banana plants are sensitivity to soil moisture stress is reflected in changes in reduced growth through reduced stomatal conductance and leaf size (Kallarackal et al., 1990) increased leaf senescence (Turner, 1998). Bananas (*Musa* spp.) rarely attain their full genetic potential for yield due to limitations imposed by water ultimately limiting the plants photosynthesis. Turner and Thomas (1998) reported that, the banana is sensitive to soil water deficits, expanding tissues such as emerging leaves and growing fruit are among the first to be affected. As soil begins to dry, stomata close and leaves remain highly hydrated, probably through root pressure. Productivity is affected because of the early closure of

stomata. Turner and Thomas (1998) who showed measurements of leaf water potential using either the exuding xylem or relative leaf water content could not be reliably linked to plant functions such as stomatal movement, net photosynthesis or leaf folding. Water potential measured by the exuding latex method appeared the best for determining leaf water status, but even this shows a small change in plants experiencing soil water deficit (Thomas and Turner, 1998) supporting the hydrated status of banana leaves although the soil is dry. Understanding banana plant response to soil moisture deficit and expression of physiological, biochemical traits are of basic scientific interest and have potential application bananas (*Musa spp*). With a view to

Treatments	3 rd MAP	5 th MAP	7 th MAP	9 th MAP	12 th MAP and Harvest	Mean
Main plot						
M ₁	101.1	160.6	228.1	246.7	251.8	197.67
M ₂	79.6	139.0	204.8	223.5	228.6	175.10
Mean	90.34	149.80	216.47	235.09	240.22	186.38
SEd	0.71	1.07	1.30	1.39	1.64	
CD (P= 0.05)	3.06	4.62	5.62	5.99	7.06	
Sub plot						
S1	129.7	217.7	334.7	383.7	431.7	299.48
S ₂	128.2	214.7	284.7	322.2	322.7	254.48
S₃	120.7	198.7	236.7	247.7	250.7	210.88
S ₄	95.7	170.7	232.7	239.7	240.7	195.88
S₅	84.8	126.8	189.8	197.8	198.8	159.61
S ₆	84.8	141.8	200.8	211.8	213.8	170.61
S ₇	78.8	172.8	231.8	243.8	245.8	194.61
S ₈	78.8	110.8	191.8	206.8	207.8	159.21
S ₉	74.2	120.2	178.7	189.7	190.7	150.66
S ₁₀	73.2	90.2	135.7	167.7	168.7	127.06
S ₁₁	72.2	105.2	187.7	201.7	201.7	153.66
S ₁₂	63.2	128.2	192.7	208.7	209.7	160.46
Mean	90.34	149.80	216.47	235.09	240.22	186.38
SEd	1.43	2.37	3.07	3.33	3.80	
CD (P= 0.05)	2.89	4.78	6.18	6.73	7.67	

Table 1. Effect of water stress on plant height (cm) at different growth stages of banana cultivars and hybrids.

Interaction effect

Treatments	3 rd MAP	5 th MAP	7 th MAP	9 th MAP	12 th MAP and Harvest	Mean
M_1S_1	136.0	224.0	341.0	390.0	438.0	305.80
M_1S_2	134.5	221.0	291.0	328.5	329.0	260.80
M_1S_3	127.0	205.0	243.0	254.0	257.0	217.20
M_1S_4	102.0	177.0	239.0	246.0	247.0	202.20
M_1S_5	96.0	138.0	201.0	209.0	210.0	170.80
M₁S ₆	96.0	153.0	212.0	223.0	225.0	181.80
M_1S_7	90.0	184.0	243.0	255.0	257.0	205.80
M₁S ₈	90.0	122.0	203.0	218.0	219.0	170.40
M₁S ₉	89.0	135.0	196.0	207.0	208.0	167.00
M_1S_{10}	88.0	105.0	153.0	185.0	186.0	143.40
M_1S_{11}	87.0	120.0	205.0	219.0	219.0	170.00
M_1S_{12}	78.0	143.0	210.0	226.0	227.0	176.80
M_2S_1	123.4	211.4	328.4	377.4	425.4	293.15
M_2S_2	121.9	208.4	278.4	315.9	316.4	248.15
M_2S_3	114.4	192.4	230.4	241.4	244.4	204.55
M_2S_4	89.4	164.4	226.4	233.4	234.4	189.55
M_2S_5	73.6	115.6	178.6	186.6	187.6	148.43
M_2S_6	73.6	130.6	189.6	200.6	202.6	159.43
M_2S_7	67.6	161.6	220.6	232.6	234.6	183.43
M_2S_8	67.6	99.6	180.6	195.6	196.6	148.03
M_2S_9	59.3	105.3	161.3	172.3	173.3	134.32
M_2S_{10}	58.3	75.3	118.3	150.3	151.3	110.72

M ₂ S ₁₁	57.3	90.3	170.3	184.3	184.3	137.32	
M_2S_{12}	48.3	113.3	175.3	191.3	192.3	144.12	
Mean	90.34	149.80	216.47	235.09	240.22	186.38	
SEd							
M at S	2.06	3.38	4.35	4.73	5.41		
S at M	2.02	3.35	4.34	4.72	5.38		
CD (P= 0.05)							
M at S	4.72	7.60	9.67	10.47	12.04		
S at M	4.09	6.76	8.75	9.51	10.85		
							-

Table 1. Cont.

elicit information on these aspects, field and laboratory investigations were undertaken.

MATERIALS AND METHODS

The experiment was carried out at national research centre for banana, Thiruchirapalli, during 2011-2012. The experiment consists of two treatments as considered as main plot and twelve cultivars and hybrids as taken as sub plots were laid out in split plot design with three replications. The main plots are, M₁ (control) with the soil pressure maintained from -0.69 to -6.00 bar, M₂ (water deficit) with the Soil pressure maintained from -0.69 to -14.00 bar. Soil pressure of -14.00 bar was reached at 30 days and measured by using soil moisture release curve and measured the soil moisture by using the pressure plate membrane apparatus (figure 1) .The sub plots are, S1: Karpuravalli (ABB), S2: Karpuravalli x Pisang Jajee, S₃: Saba (ABB), S₄: Sanna Chenkathali (AA), S₅: Poovan (AAB), S₆: Ney poovan (AB), S₇: Anaikomban (AA), S₈: Matti x Cultivar Rose, S₉: Matti (AA), S₁₀: Pisang Jajee x Matti, S₁₁: Matti x Anaikomban and S₁₂: Anaikomban x Pisang Jajee. The morphological characters viz., plant height and number of leaves were measured at 3rd, 5th, 7th, 9th month after planting and at harvest stages of the crop and physiological components like, Relative water content by the method of Weatherly (1950) and expressed in percentage, Total chlorophyll were estimated in physiologically active leaves as per the procedure of Hixcox and Israelstam (1979) and expressed as mg g fresh weight and Osmotic potential were recorded by using the Vapor Pressure Osmometer (VAPRO 5520 meter) during 3rd, 5th, 7th, 9th month after planting and at harvest stages of the crop. The yield and yield components were assessed at the time of harvesting.

RESULT AND DISCUSSION

Plant height

The time trend of plant height of banana cultivars revealed

a progressive increase from 3rd MAP to harvest stage (Table 1). Comparison of two treatments at main plot level revealed that M₁ recorded significantly higher plant height than M_2 . Among the sub plot treatments, S_1 observed to be the tallest plant with the height of 431.7cm followed by S_2 (322.7cm) and S_3 (250.7cm). The other sub plot treatments like S₄, S₆, S₇, S₈, S₁₁ and S₁₂ showed medium height in the range of 201.7 to 245.8cm and S_5 , S_9 and S_{10} found to be the dwarfed plants with height ranging from 168.7 to 198.8cm at harvest stage. The interaction effects of M at S and S at M were significant at all growth stages. Among the interaction treatments, M₁S₁ recorded taller plants of 438.0cm. This was very closely followed by M_1S_2 (329.0cm). The main plot M₂ resulted in 3 to 5 per cent reduction in height of the plants of M_2S_1 , M_2S_2 , M_2S_3 and M_2S_4 over the treatments of M_1S_1 , M_1S_2 , M_1S_3 and M_1S_4 at 7th MAP. The other treatments, M_2S_5 , M_2S_6 , M_2S_7 and M_2S_8 showed 9 to 11 per cent reduction over M₁S₅, M₁S₆, M₁S₇ and M₁S₈. The treatments of M_2S_9 , M_2S_{10} , M_2S_{11} and M_2S_{12} showed greater reduction in height of the plant with the range of 16-22 per cent over the subplot interaction with M1 at 7th MAP stage.

Plant height is an important morphological parameter related to growth and development of the crop. Growth involves both cell growth and development. Cell growth and development is a process consisting of cell division, cell enlargement and cell differentiation (Wareing and Phillips, 1970). These processes are very sensitive to water deficit because of their dependence upon turgor. Morphologically, plant growth is perceived as an increase in plant size in terms of plant height and growth rate, while development involves tissue and organ formation. The influence of water deficit on the growth of banana cultivars exhibited significant variations at all the growth stages. Application of irrigation at 80% available soil moisture caused a significant improvement in plant height, irrigation at 50% available soil moisture resulted in a considerable reduction in plant height. The most evident effect of water deficit to the plant growth of banana was growth inhibition. Cell expansion and enlargement is one of the most sensitive processes affected by a change in plant water status (Begg and aff-

Treatments	3 rd MAP	5 th MAP	7 th MAP	9 th MAP	Harvest	Mean
Main plot						
M ₁	10.0	10.3	11.6	10.0	8.8	10.16
M ₂	8.5	8.8	10.2	8.5	7.3	8.68
Mean	9.25	9.58	10.88	9.29	8.08	9.42
SEd	0.062	0.060	0.071	0.067	0.048	
CD (P= 0.05)	0.269	0.259	0.308	0.290	0.207	
Sub plot						
S1	12.5	12.5	14.5	14.0	10.5	12.80
S ₂	11.5	11.5	12.5	10.5	9.5	11.10
S ₃	11.5	11.5	12.5	10.5	9.5	11.10
S ₄	9.5	10.5	11.5	10.5	8.5	10.10
S_5	9.3	9.3	11.3	9.3	8.3	9.45
S_6	9.3	8.3	10.3	8.3	7.3	8.66
S ₇	9.3	9.3	10.3	9.3	8.3	9.25
S ₈	8.3	9.3	10.3	9.3	8.3	9.05
S ₉	8.0	8.0	9.0	7.0	7.0	7.80
S ₁₀	8.0	9.0	10.0	8.0	7.0	8.40
S ₁₁	7.0	9.0	10.0	7.0	6.0	7.80
S ₁₂	7.0	7.0	9.0	8.0	7.0	7.60
Mean	9.25	9.58	10.88	9.29	8.08	9.42
SEd	0.133	0.139	0.150	0.145	0.113	
CD (P= 0.05)	0.269	0.281	0.302	0.292	0.227	

Table 2. Effect of water stress on number of leaves at different growth stages of banana cultivars and hybrids.

Interaction effect

Treatments	3 rd MAP	5 th MAP	7 th MAP	9 th MAP	Harvest	Mean
M_1S_1	13.0	13.0	15.0	14.5	11.0	13.30
M_1S_2	12.0	12.0	13.0	11.0	10.0	11.60
M_1S_3	12.0	12.0	13.0	11.0	10.0	11.60
M_1S_4	10.0	11.0	12.0	11.0	9.0	10.60
M_1S_5	10.0	10.0	12.0	10.0	9.0	10.20
M_1S_6	10.0	9.0	10.0	9.0	8.0	9.20
M ₁ S ₇	10.0	10.0	11.0	10.0	9.0	10.00
M_1S_8	9.0	10.0	11.0	10.0	9.0	9.80
M_1S_9	9.0	9.0	10.0	8.0	8.0	8.80
M ₁ S ₁₀	9.0	10.0	11.0	9.0	8.0	9.40
M ₁ S ₁₁	8.0	10.0	11.0	8.0	7.0	8.80
M ₁ S ₁₂	8.0	8.0	10.0	9.0	8.0	8.60
M_2S_1	12.0	12.0	14.0	13.5	10.0	12.30
M_2S_2	11.0	11.0	12.0	10.0	9.0	10.60
M_2S_3	11.0	11.0	12.0	10.0	9.0	10.60
M_2S_4	9.0	10.0	11.0	10.0	8.0	9.60
M_2S_5	8.5	8.5	10.5	8.5	7.5	8.70
M_2S_6	8.5	7.5	9.5	7.5	6.5	7.90
M_2S_7	8.5	8.5	9.5	8.5	7.5	8.50
M_2S_8	7.5	8.5	9.5	8.5	7.5	8.30
M_2S_9	7.0	7.0	8.0	6.0	6.0	6.80

M ₂ S ₁₀	7.0	8.0	9.0	7.0	6.0	7.40
M_2S_{11}	6.0	8.0	9.0	6.0	5.0	6.80
M_2S_{12}	6.0	6.0	8.0	7.0	6.0	6.60
Mean	9.25	9.58	10.88	9.29	8.08	9.42
SEd						
M at S	0.191	0.198	0.215	0.207	0.160	
S at M	0.189	0.197	0.212	0.205	0.159	
CD (P= 0.05)						
M at S	0.433	0.441	0.489	0.468	0.356	
S at M	0.381	0.397	0.428	0.413	0.322	

ected by a change in plant water status (Begg and Turner, 1976).

Number of leaves

The leaf production in terms of number of leaves per plant revealed an increasing trend from 3rd MAP to 7th MAP with a decline thereafter (Table 2). Comparing the main plot treatments, M_1 recorded significantly higher leaf number (11.6) at 7th MAP than M_2 , which recorded the mean leaf number of 10.2 indicating a considerable reduction (14%) over M1. The leaf production was observed to be high at 7th MAP in S₁ (14.5 No plant¹) followed by S_2 (12.5 No. plant⁻¹) and S_3 (12.5 No. plant⁻¹). The lowest number of 9.0 leaves was, however, produced by S₉ and S₁₂. The interaction effects of M at S and S at M were significant at all growth stages. The treatment M_1S_1 performed better than other treatments with the production of higher leaf number (15.0) at 7th MAP. The interaction between M_2 and subplots showed a considerable reduction in leaf production over the interaction between M1 and subplot treatments. The interaction treatment M₂S₁, M₂S₂, M₂S₃ and M₂S₄ showed a reduction of about 7 to 8 per cent, whereas $M_2S_5 M_2S_6$. M_2S_7 and M_2S_8 exhibited a reduction ranging from 11 to 14 per cent. The other treatments M_2S_9 , M_2S_{10} , M_2S_{11} and M_2S_{12} however, showed greater reduction of 18 to 21 percent in leaf production over the interaction between M1 and sub plot treatments.

Leaf is considered as an important functional unit of plant, it is the most effective weapons in the crop economy and eventually their yield under the drought stress. The increase in **number of leaves** represents an increase in the photosynthetic surface of the plant. In the present study, a significant increase was observed in number of leaves in plants under irrigation at 80% available soil moisture compared to those under irrigation at 50% available soil moisture. Turner and Thomas (1998) suggested that leaf folding may reflect leaf water status and soil water deficit leads to arresting of newer leaf production followed by decreased number of leaves in banana. The banana plant is very sensitive to water deficiency and this is reflected by reduced greenness of foliage. When the water deficiency become severe all the leaves fall prematurely and the pseudostem tissue collapses at a point about mid way between the ground and lowest leaves and the plant falls over (Stover and simmonds, 1987). The reduction in leaf number under water stress may have been due to reduction in leaf formation and increased abscission of lower leaves eventually leading to wilting of the whole plant (Tezara *et al.*, 2002). According to Turner (1993), the reduction in leaf number under severe water deficit was partially due to leaf senescence. Hsiao (1973), stated that growth inhibition after wilting of leaves and plant is known to enhance nucleic acid destruction.

Relative Water Content (RWC: %)

The data on RWC revealed a progressive increase from 3rd to 7Th MAP with a decline thereafter. The main and sub-plots treatments differed significantly at all the growth stages. The treatment M1 outperformed with better RWC value of 78.2 per cent at 7th MAP stage, whereas M₂ recorded significantly lesser RWC value of 68.7%. Among the sub-plot treatment varieties, S_1 was found to be effective in maintaining higher RWC value (83.8%) over S_{12} (63.7%), which was followed by S_2 (83.1%) and S₃ (80.2%). All the interaction treatments registered significant differences at all the stages, therefore, M at S and S at M attained differences significantly. Treatment M₁S₁ registered higher RWC of 86.1 percentage followed by M_1S_2 (85.4%), M_1S_3 (82.5%) and M_1S_4 (82.0%). However, a considerable reduction could also be noticed in RWC due to interaction with M₂ and subplot treatments. M_2S_1 , M_2S_2 , M_2S_3 and M_2S_4 maintained its superiority (81.5, 80.8, 77.9 and 77.4 per cent) with about 5 to 8 per cent reduction, whereas, all the other treatments showed about 12 to 20 per cent reduction than M₁ and subplot treatments.

Relative Water Content (RWC) is the appropriate measure of plant water status in terms of the physiological consequence of cellular water deficit. It was



Figure 2. Effect of water stress on total chlorophyll content (mg g⁻¹) of banana cultivars and hybrids at different growth stages.

used instead of plant water potential as RWC referring to its relation with cell volume, which could accurately indicate the balance between absorbed water by plant and lost through transpiration. The banana plants are able to maintain their internal water status during drought by reducing radiation load and closing stomata (Thomas and Turner, 1998). The RWC was estimated in order to find out the plant water status of banana cultivars under water stress situations. Leaf RWC had a significant influence on photosynthesis, by reducing the net photosynthesis by more than 50 per cent when RWC was less than 80 per cent. As observed by Slatver (1955), a reduction by 5% in RWC led to reduction in photosynthesis by 40 to 50 %. The early reduction of stomatal conductance and the minor diminution of leaf RWC could indicate that the banana plants showed a drought avoidance mechanism to maintain a favorable plant water status involving stomatal closure in response

to water stress (Turner and Lahav 1983).

Total chlorophyll content (mg g⁻¹)

The data on total chlorophyll content reflected similar time trend of chlorophyll 'a', chlorophyll 'b' (Figure 2). Main plot treatments differed significantly at all growth stages. Among the main plot treatments, M_1 out performed with higher total chlorophyll content of 1.03 mg g⁻¹ than M_2 (0.90 mg g⁻¹) showing a 13 per cent reduction over M_1 at 7th MAP stage. With regard to the sub-plot treatment S₁ recorded higher total chlorophyll content of 1.31 mg g⁻¹ closely followed by S₂ (1.17 mg g⁻¹) and S₃ (1.08 mg g⁻¹). S₁₂ however, recorded the lowest content of 0.72 mg g⁻¹ among the subplot treatments at 7th MAP stage. Significant differences among the interaction treatments also revealed the differential responses of M₁

Treatments	3 rd MAP	5 th MAP	7 th MAP	9 th MAP	Harvest	Mean
Main plot						
M ₁	0.37	0.69	0.53	0.71	0.63	0.58
M ₂	0.42	0.73	0.72	0.90	0.81	0.72
Mean	0.39	0.71	0.63	0.80	0.72	0.65
SEd	0.003	0.005	0.007	0.011	0.008	
CD (P= 0.05)	0.015	0.024	0.034	0.050	0.037	
Sub plot						
S ₁	0.49	0.86	0.75	0.92	0.82	0.77
S ₂	0.36	0.75	0.67	0.85	0.75	0.68
S ₃	0.38	0.75	0.58	0.86	0.76	0.66
S ₄	0.36	0.75	0.56	0.93	0.83	0.68
S ₅	0.43	0.79	0.64	0.80	0.70	0.67
S ₆	0.42	0.80	0.62	0.81	0.76	0.68
S ₇	0.35	0.69	0.64	0.83	0.77	0.65
S ₈	0.39	0.67	0.62	0.80	0.73	0.64
S ₉	0.37	0.66	0.63	0.74	0.67	0.62
S ₁₀	0.39	0.63	0.61	0.72	0.65	0.60
S ₁₁	0.43	0.61	0.68	0.68	0.62	0.60
S ₁₂	0.37	0.58	0.53	0.71	0.58	0.55
Mean	0.39	0.71	0.63	0.80	0.72	0.65
SEd	0.004	0.006	0.008	0.009	0.007	
CD (P= 0.05)	0.009	0.013	0.017	0.018	0.014	

Table 3. Effect of water stress on leaf Osmotic Potential (MPa) at different growth stages of banana cultivars and hybrids.

Interaction effect

Treatments	3 rd MAP	5 th MAP	7 th MAP	9 th MAP	Harvest	Mean
M ₁ S ₁	0.43	0.77	0.62	0.79	0.69	0.66
M_1S_2	0.30	0.70	0.54	0.72	0.62	0.58
M_1S_3	0.32	0.68	0.45	0.73	0.63	0.56
M_1S_4	0.33	0.69	0.43	0.80	0.70	0.59
M₁S₅	0.39	0.74	0.55	0.71	0.61	0.60
M ₁ S ₆	0.41	0.75	0.53	0.72	0.67	0.62
M ₁ S ₇	0.28	0.72	0.55	0.74	0.68	0.59
M ₁ S ₈	0.34	0.69	0.53	0.71	0.64	0.58
M ₁ S ₉	0.35	0.67	0.57	0.68	0.61	0.58
M_1S_{10}	0.43	0.64	0.55	0.66	0.59	0.57
M ₁ S ₁₁	0.48	0.57	0.62	0.62	0.56	0.57
M_1S_{12}	0.33	0.61	0.47	0.65	0.52	0.52
M_2S_1	0.55	0.95	0.88	0.99	0.95	0.88
M_2S_2	0.41	0.80	0.80	0.98	0.88	0.78
M_2S_3	0.43	0.81	0.71	0.99	0.89	0.77
M_2S_4	0.38	0.80	0.69	0.99	0.96	0.78
M_2S_5	0.47	0.83	0.73	0.89	0.79	0.74
M_2S_6	0.43	0.84	0.71	0.90	0.85	0.74
M_2S_7	0.42	0.66	0.73	0.92	0.86	0.72
M_2S_8	0.44	0.64	0.71	0.89	0.82	0.70
M_2S_9	0.39	0.65	0.70	0.81	0.74	0.66
M_2S_{10}	0.34	0.61	0.68	0.79	0.72	0.63
M_2S_{11}	0.38	0.64	0.75	0.75	0.69	0.64
M_2S_{12}	0.40	0.55	0.60	0.78	0.65	0.59
Mean	0.39	0.71	0.63	0.80	0.72	0.65

Table 3. cont.						
SEd						
M at S	0.007	0.013	0.014	0.014	0.013	
S at M	0.006	0.012	0.013	0.009	0.010	
CD (P= 0.05)						
M at S	0.018	0.032	0.039	0.051	0.039	
S at M	0.013	0.024	0.026	0.019	0.020	

Correlation studies graph.





M₁S₁ registered higher total chlorophyll content of 1.35

and M_2 treatments over sub-plot treatments. Among them

mg g⁻¹ over M₂. Treatments such as M_1S_2 and M_1S_3 also performed better than other treatments with 7 to 13 per cent increase over M₂ treatment interaction. A considerable reduction in total chlorophyll content could also be observed due to interaction with M₂, the percentage however, varies with different sub plots. Among the sub plot treatments, M₂S₁, M₂S₂, M₂S₃ and M₂S₄ exhibited 6 to 12 per cent reduction in total chlorophyll content, whereas, all the other treatments showed 13 to 17 per cent reduction over M₁ and subplot interaction.

The chloroplast in green plants constitutes the photosynthetic apparatus. Chlorophylls and other photosynthetic pigments are found in the form of protein pigment complexes mainly in thylakoid membranes of grana. Photosynthetic pigments play major role in plant productivity, as they are responsible for capturing light energy and using it as a driving force for producing the assimilates. Water deficit induces disintegration of thylakoid membranes and causes degradation of chlorophyll pigments. This could substantially contribute to the overall inhibition of photosynthesis in leaves of water deficit plants (Farquhart et al., 1982). The mechanism of reduction in chlorophyll content due to the enhancement of chlorophyllase activity in water stressed plants could be the cause for chlorophyll degradation. Ghavami (1973) noticed a drastic reduction in the total chlorophyll content under water deficit condition due to the disruption of fine structure of chloroplast and instability of pigment and enhanced chlorophyllase activity. Thomas and Turner (2001) also observed a decrease in chlorophyll content in banana cultivars leading to decrease in photosynthesis.

Osmotic potential (MPa)

The data on osmotic potential revealed a progressive increase upto 9th MAP and declined at harvest. The main and sub-plot treatments differed significantly at all the growth stages (Table 3). Between the two main plot treatments, M₂ had significantly higher osmotic potential (0.42, 0.73, 0.72, 0.90 and 0.81 MPa) over M₁ at 3rd, 5th, 7th, 9th and at harvest respectively. All the sub-plot treatments also significantly differed. Among the sub- plot treatments S₁ recorded the higher osmotic potential of 0.49, 0.86, 0.75, 0.92, and 0.82 MPa at 3rd, 5th, 7th, 9th and harvest stage respectively. This treatment was followed by S₂, S₃ and S₄. The lowest osmotic potential was registered by S₁₂ (0.53 MPa). The interaction effects of M at S and S at M revealed significant difference at all the stages of growth. At 9th MAP, M_2S_1 , M_2S_2 , M_2S_3 and M_2S_4 maintained a high osmotic potential of 0.99 MPa, whereas the treatments M_2S_{10} , M_2S_{11} and M_2S_{12} recorded the osmotic potential of around 75 to 79 per cent.

Osmotic potential is considered as an important physiological mechanism of drought adaptation in banana plants (Turner, 1972). Osmotic Adjustment requires regulation of intracellular levels of several compounds

collectively known as osmolytes. In banana, the osmotic potential was determined from xylem sap. Kallarackal et al. (1986) stated that the solute potential of exuding latex provided an excellent guide to the water status of the plant during water deficit conditions. Banana latex contains large number of vacuolysosomal organelles called "lutoids" which are capable of actively transporting ions across the membranes with higher osmotic potential activity during stress conditions. In this present investigation, the cultivars like Karpuravalli, Karpuravalli x Pisang jajee, Saba and Sannachenkathali registered higher osmotic potential with 50 per cent increase over control at 7th MAP. It can be concluded that, higher osmotic adjustment under stress conditions is considered as important physiological mechanism of drought an adaptation in many plants (Subbarao et al., 2000). The cultivars like Matti, Matti x Anaikomban, Matti x cultivar rose and Pisang jajee x Matti showed only 20 per cent increase in osmotic potential than the control. It was also established that the increase in osmotic potential in response to water stress is a behavior which causes a more water stress tolerance (Turner and Lahav 1983).

Correlation studies with yield

The final yield of crop is the cumulative effects of growth attributes and such of those treatments which manipulate the favourable parameters could result in the positive relationship with higher productivity. The relationships of number of leaves, total chlorophyll content and Relative Water Content were correlated with the final yield presented in the Fig 3, Fig 4 and Fig 5 at 7th MAP. Based on the results arrived from the correlation revealed that the correlation between number of leaves, total chlorophyll content were correlated water Content were correlated water content were number of leaves, total chlorophyll content and Relative Water Content were correlated were showed significant positive correlation with yield.

REFERENCES

- Begg, J.E and N.C. Turner. 1976. Crop water deficits. Adv. Agron., 28: 161-217.
- Bray EA, Bailey-Serres J, Weretilnyk E (2000). Responses to abiotic stresses. In Biochemistry and Molecular Biology of Plants, B.B. Buchanan, W. Gruissem, and R.L. Jones, eds (Rockville, MD: American Society of Plant Physiologists), pp. 1158–1203.
- Cambridge Univ. Press. Cambridge, pp. 47-67.
- Diethelm R, R Shibles (1989). Relationship of enhanced sink demand with photosynthesis and amount and activity of Ribulose-1,5 biphosphate carboxylase in soybean leaves. J. Plant Physiol., 134: 70-74.
- Farquhart GD, Wong SO, Evans TR, Hubiok KT (1982). Photosynthesis and gas exchange. In H.G.Jones, T.

- Ghavami M (1973). Determining water needs of the banana plant. Transactions of the American Society of Agricultural Engineers 16:598-600.
- Hsiao JC, Acevedo E (1974). Plant responses to water deficit and drought resistance. Agric. Meteorol., 14: 59-84.
- Hsiao TC (1973). Plant responses to water deficit. Ann. Rev. Plant Physiol. 24:519:570.

J. Flowers, and M.B.Jones (eds.), Plant Under Stress.

- Joseph, MC, Randall DD, Nelson CJ (1981). Photosynthesis in polyploid tall fescue. II. Photosynthesis and RUBP case of polyploid tall fescue. Plant Physiol., 68: 894-898.
- Kallarackal J, Milburn JA, Baker DA (1990). Water relations of the banana. III effects of controlled water stress on water potential, transpiration, photosynthesis and leaf growth. Aust. J. Plant Physiol. (17) : 79-90.
- Kallarackal J, Milburn JA, Baker DA (1986). Transpiration characteristics of banana leaves inresponse to progressive depletion of available soil moisture. Sci. Hortic. 30: 289-300.
- Martignone RA, Guiamot JJ, Nakayama F (1987). Nitrogen partitioning and leaf senescence in soybean as related to nitrogen supply. Field Crops Res., **17**: 17-20.
- Pallar, JE, Stansel JR, Koske M (1979). Effect of drought on flour runner peanut. Agron. J., 71: 853-858.
- Robinson JC (1984). Bananas and Plantains. Wallingford, CAB International.
- Slatyer RO (1955). Aust. J. Agric. Res., 61: 365-377.
- Stover RH, Simmonds NW (1987). Bananas (3rd edition) Longman, London, United Kingdom.
- Tezara W, Mitchell V, Driscoll SP, Lawlor DW (2002).

Effects of water deficits and its interaction with CO₂ supply on the biochemistry and physiology of photosynthesis in sunflower. J. Expt. Bot. 53, 1781-1791.

- Thomas DS, Turner DW (2001). Banana leaf gas exchange and chlorophyll fluorescence in response to soil drought, shading and lamina folding. Scientia horticulturae 90:93-108.
- Turner DW (1972). Banana plant growth. 1. Groeth. 1. Gross morphology. Aust, J. Expt, Agric. Animal Husband .12:216-224.
- Turner DW (1993). Bananas and plantains. In Handbook of Environmental Physiology of Fruit Crops Volume II : Sub-tropical and Tropical Crops, 37-64 (Eds. B. Schaffer and P. C. Andersen). Boca Raton, CRC Press.
- Turner DW (1998). The impact of environmental factors on the development and productivity of bananas and plantains. In Proceedings of the 13th ACORBAT meeting, Guayaquil, Ecuador, 635-663 (Ed. L. H. Arizaga). Ecuador, CONABAN.
- Turner DW, Lahav E (1983). The growth of banana plants in relation to temperature. Aust. J. Plant Physiol. 10:43-53.
- Turner DW, Thomas DS (1998). Measurements of plant and soil water status and their association with leaf gas exchange in banana (Musa spp.): a laticiferous plant. Scientia Horticulturae 77:177-193.
- Wahad Z, Zulkifli Hj, Suzita S (2000). Proceedings of the first national banana seminar at Awana Genting and country resort. UPM: Serdang (mys). 343 p.
- Wareing PF, Phillips IDJ (1970). The control and development in plants. Pergamon Press, Oxford. 1-21.